

Abstract

Regulatory haplotypes modulate the expressivity and penetrance of hereditary cancer-risk mutations. We demonstrate expression quantitative trait loci (eQTL) detection using Ambry CancerNext and RNAInsight data from 57,898 patients, one of the largest clinical blood paired DNA-RNA datasets, validated against GTEx v8 whole blood eQTL data. After batch renormalization, 66 eQTLs were identified, showing strong additive effects ($R^2 = 0.99$, $p < 1e-16$). Select eQTLs correlated with age of onset in patients with loss-of-function mutations and variants of uncertain significance, even after adjusting for confounders. These findings support eQTL detection in clinical datasets and suggest regulatory haplotypes may refine risk assessment in hereditary cancer.

Introduction

Inference of regulatory haplotypes which modify expressivity and penetrance of coding hereditary cancer-risk mutations is a key goal of precision medicine, potentially enabling deeper insight into individual risk profiles and therapeutic responses. Previous seminal work has already established that powerful functional readouts of latent regulatory variants acting on genes using phased haplotypes of coding variants and regulatory variants, including eQTL mapping in cis, can elucidate the enrichment of penetrance increasing configurations for pathogenic variants. In this work we demonstrate the feasibility of eQTL detection using Ambry CancerNext and RNAInsight data from 57,898 matched patients and present a preliminary eQTL analytic association analysis for potential risk refinement.

Methods and Materials

- Ambry CancerNext and RNAInsight data from 57,898 samples were analyzed using genetic test results and matched gene expression measurements (TPM) respectively.
- GTEx v8 (whole blood) was used as a gold standard set to test for eQTL candidates in the Ambry data.
- eQTL concordance was defined as:
 - significant expression change between wildtype and heterozygous alternative allele with Bonferroni adjusted p-value < 0.05 , and
 - having the same direction of effect changes between Ambry and GTEx data.
 - Effect size was calculated by batch-corrected TPM by accounting for sequencing run, gender, and ethnicity.
- Dosage ratio was calculated as $\frac{|WT-HOM|}{2 \times |WT-HET|}$, where WT, HET, and HOM represent the median TPM of patients with wildtype, heterozygous, and homozygous alternative alleles, respectively. We defined "Additive" as ratio within 1 sd, "HOM-Dominant" as ratio $> 1 + sd$, and "WT-Dominant" as ratio $< 1 - sd$, where sd is the standard deviation (sd) of the 66 concordant eQTLs.
- Association and covariate renormalization analyses were performed using penalized linear models. Sequencing run, gender, and ethnicity were included as covariates. Standard statistical hypothesis testing methods were applied.

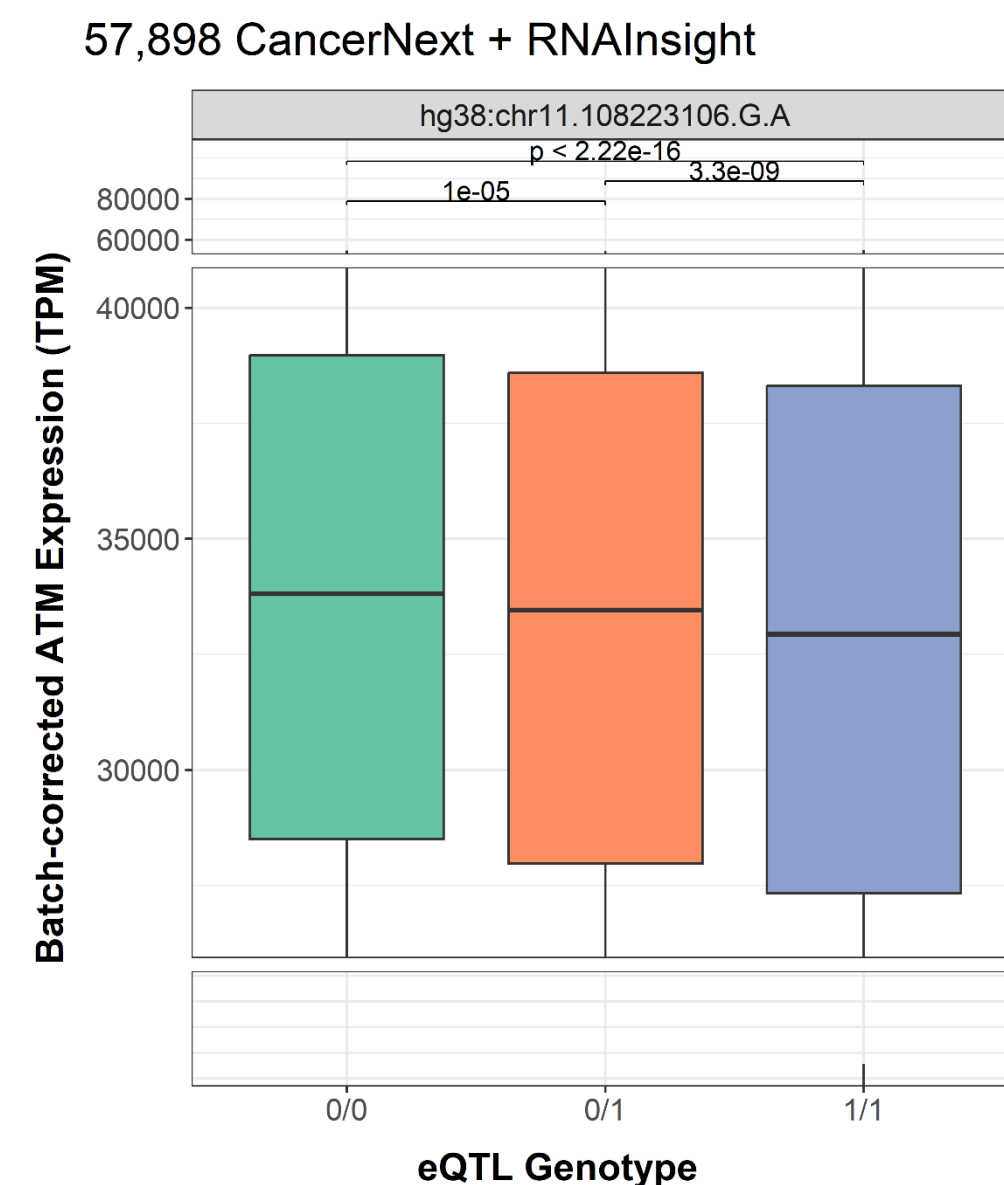


Figure 1. Example of a significant eQTL in ATM gene from Ambry CancerNext + RNAInsight data. The ATM batch-corrected expression decreased with each additional copy of the common SNP, which is consistent with the understanding that mutations in ATM can significantly increase the risk of developing breast cancer (BrCa).

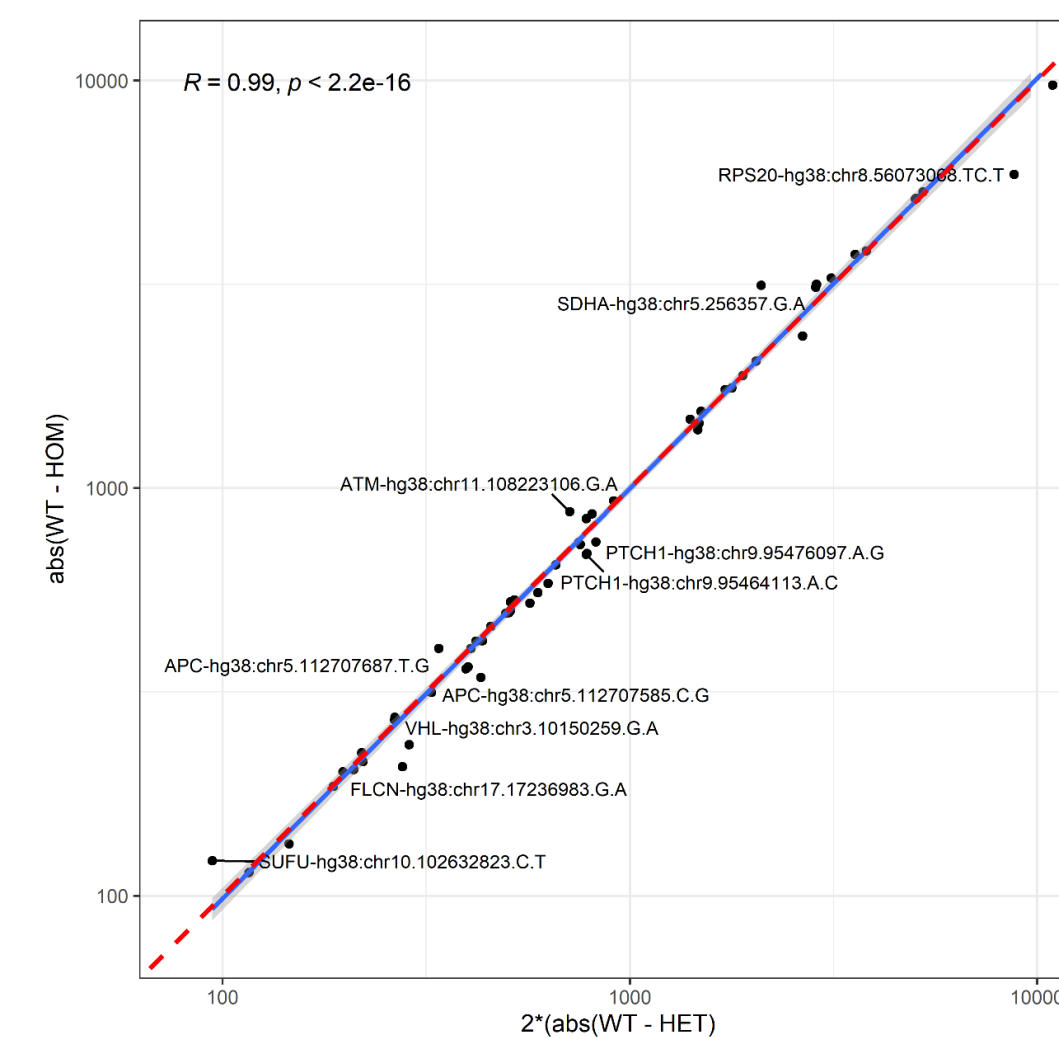


Figure 2. Dosage plot from 66 Ambry eQTLs that are concordant with GTEx whole blood data. The X-axis represents twice of the effect size between wildtype and heterozygous variant, and the Y-axis represents the effect size between wildtype and homozygous variant. Non-additive eQTLs are labeled by the gene names and hg38 genomic coordinates.

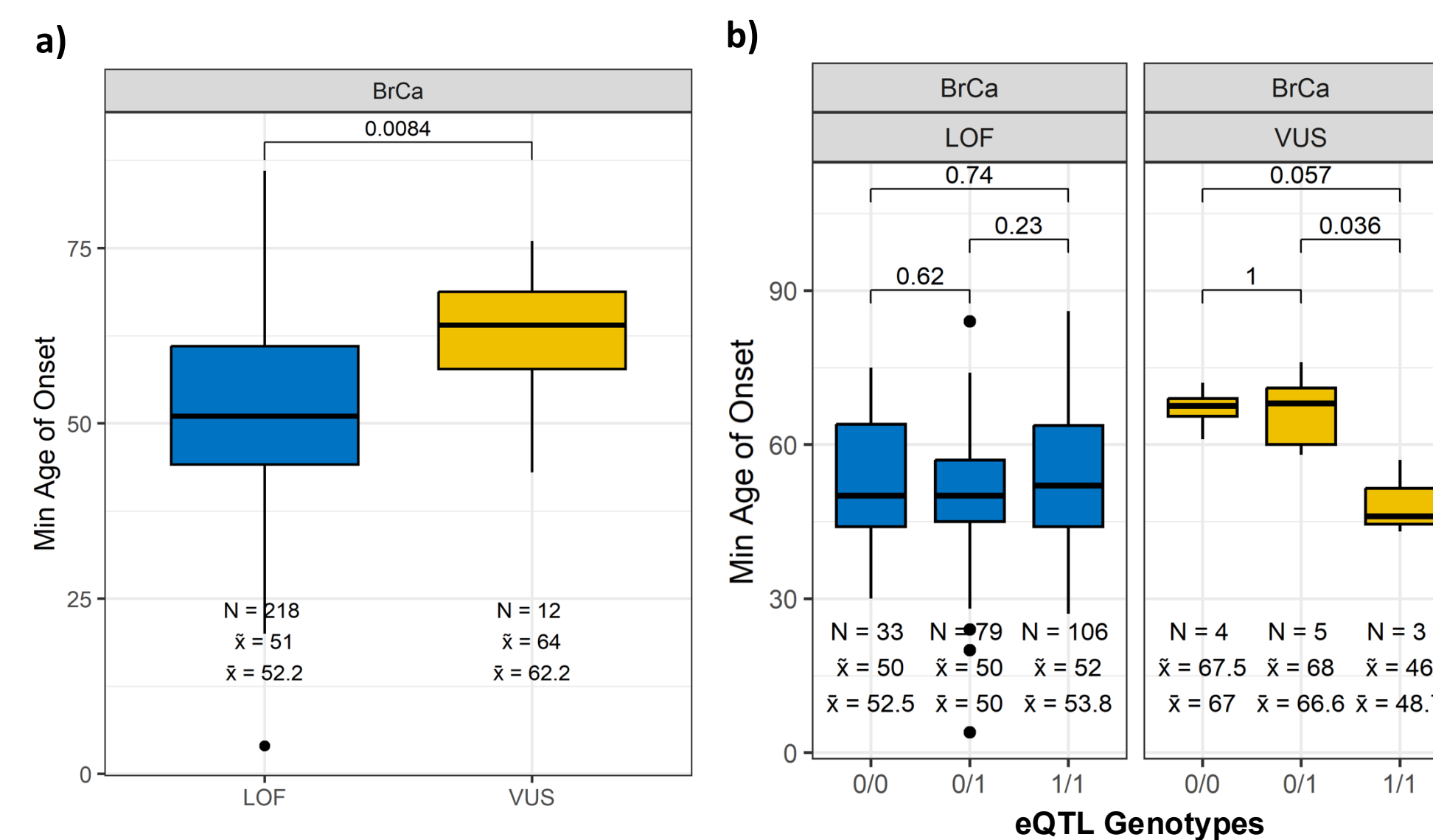


Figure 3. a) Age of onset comparison in Ambry BrCa patients from 57,898 CancerNext + RNAInsight data with VHL loss of function (LOF) variants or variants of unknown significance (VUS); b) same data as in a) but grouped the samples by the VHL eQTL genotypes (hg38:chr3.10150259.G>A). LOF: Samples with LOF variants in VHL and high-penetrance genes, such as BRCA1, BRCA2, TP53, PALB2, or CHEK2; VUS: Samples with any VUS variants in VHL.

Results

- Ambry CancerNext+RNAInsight data from blood enabled eQTL detection. An example of significant eQTL in ATM is shown in **Figure 1**. The direction of expression change is consistent with our understanding that mutations in the ATM can significantly increase risk of developing breast cancer (BrCa).
- We observed a strongly additive dosage pattern ($R^2 = 0.99$, $p < 1e-16$) from 66 out of 96 eQTLs concordant with GTEx whole blood data after batch correction (**Figure 2, Table 1**). The 96 testable eQTLs are from 24 genes.
- Using select eQTLs we showed that age of onset in patients positive with loss of function mutations and variants of uncertain significance is correlated with eQTL genotype even after accounting for possible clinical and technical confounders (**Figure 3**).

Table 1. Ambry eQTLs concordant with GTEx blood eQTLs. Red: upregulating; blue: downregulating.

Gene	Variant hg38	Group	Gene	Variant hg38	Group
SDHA	chr5.256357.G.A	HOM-Dominant	SDHA	chr5.235249.C.T	Additive
SUFU	chr10.102632823.C.T	HOM-Dominant	SMARCB1	chr22.23834293.C.CG	Additive
ATM	chr11.108223106.G.A	HOM-Dominant	MSH2	chr2.47385739.G.A	Additive
APC	chr5.112707687.T.G	HOM-Dominant	MSH2	chr2.47373967.T.C	Additive
SDHA	chr5.224518.A.G	Additive	RAD51C	chr17.58358748.C.T	Additive
SDHA	chr5.226045.A.C	Additive	KIF1B	chr1.10361820.A.G	Additive
SDHA	chr5.228247.T.C	Additive	MSH2	chr2.47385085.C.T	Additive
SDHA	chr5.251354.G.A	Additive	MSH3	chr5.80654880.TG.CAGCGGCTG.CAGCGGCT	Additive
SDHA	chr5.251426.A.G	Additive	RAD51C	chr17.58357496.C.T	Additive
SDHA	chr5.254484.A.T	Additive	ATRIP	chr3.48446788.G.C	Additive
SDHA	chr5.233619.C.G	Additive	MSH3	chr5.80654678.C.T	Additive
MSH6	chr2.47790942.A.G	Additive	SDHC	chr1.161362556.C.G	Additive
MSH6	chr2.47783419.C.A	Additive	RAD51C	chr17.58415439.T.C	Additive
MSH2	chr2.47466649.A.T	Additive	MSH6	chr2.47798625.C.T	Additive
CTNNA1	chr5.138812254.A.C	Additive	BARD1	chr2.214780821.C.G	Additive
SDHA	chr5.256394.G.A	Additive	MEN1	chr11.64810148.G.C	Additive
BARD1	chr2.214780740.C.G	Additive	KIF1B	chr1.10378629.C.T	Additive
MSH2	chr2.47386731.T.C	Additive	SDHC	chr1.161314434.A.AGT	Additive
MSH3	chr5.80854162.A.G	Additive	ATM	chr11.108270974.G.T	Additive
MLH3	chr14.75017109.T.C	Additive	MLH1	chr3.36993455.G.A	Additive
MLH3	chr14.75047125.G.A	Additive	SUFU	chr10.102627262.T.C	Additive
RAD51C	chr17.58692618.C.T	Additive	LZTR1	chr22.20983036.G.A	Additive
BARD1	chr2.214809599.C.G	Additive	LZTR1	chr22.20985799.T.C	Additive
BARD1	chr2.214809617.C.G	Additive	MSH3	chr5.80654689.C.T	Additive
CTNNA1	chr5.138886300.G.C	Additive	PTCH1	chr9.95464025.C.T	Additive
CTNNA1	chr5.138930857.G.A	Additive	RPS20	chr8.56074389.T.C	Additive
CTNNA1	chr5.138934602.T.TATC	Additive	FLCN	chr17.17231866.G.A	Additive
CTNNA1	chr5.138934853.T.A	Additive	PTCH1	chr9.95466097.A.G	WT-Dominant
MSH3	chr5.80654962.A.G	Additive	PTCH1	chr9.95464113.A.C	WT-Dominant
PMS2	chr7.5987357.G.A	Additive	VHL	chr3.10150259.G.A	WT-Dominant
BARD1	chr2.214809647.C.T	Additive	APC	chr5.112707585.C.G	WT-Dominant
MSH2	chr2.47385725.C.CTG	Additive	FLCN	chr17.17236983.G.C	WT-Dominant
SMARCB1	chr22.23825326.G.A	Additive	RPS20	chr8.56073068.T.C.T	WT-Dominant

Summary & Conclusion

Our findings highlight the value of integrating regulatory variant analysis with clinical genetic testing to refine individual cancer risk profiles. The identification of eQTLs in a large clinical cohort underscores their potential to modify disease penetrance and expressivity, offering new avenues for precision medicine. These results support the use of eQTL mapping for risk refinement and suggest that germline eQTL variation may provide mechanistic prior information that complements tumor based HRD RNA signatures, enabling more refined identification of HRD-like biology beyond canonical BRCA1/2 alterations.

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